This article was downloaded by:

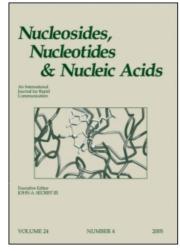
On: 27 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

Antiherpes Simplex Virus Activity of 9-[4-Hydroxy-3-(hydroxymethyl)-1-butyl] guanine

Michael A. Tippie^a; John C. Martin^a; Donald F. Smee^a; Thomas R. Matthews^a; Julien P. M. Verheyden^a Syntex Research, Palo Alto, CA

To cite this Article Tippie, Michael A. , Martin, John C. , Smee, Donald F. , Matthews, Thomas R. and Verheyden, Julien P. M.(1984) 'Antiherpes Simplex Virus Activity of 9-[4-Hydroxy-3-(hydroxymethyl)-1-butyl] guanine', Nucleosides, Nucleotides and Nucleic Acids, 3: 5, 525 — 535

To link to this Article: DOI: 10.1080/07328318408081287 URL: http://dx.doi.org/10.1080/07328318408081287

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

ANTIHERPES SIMPLEX VIRUS ACTIVITY OF 9-[4-HYDROXY-3-(HYDROXYMETHYL)-1-BUTYL]GUANINE

Michael A. Tippie, John C. Martin,* Donald F. Smee, Thomas R. Matthews, and Julien P.H. Verheyden

Syntex Research, Palo Alto, CA 94304

Abstract. The carba analogue 2 of DHPG (1) was found to be highly inhibitory to herpes simplex virus type 1 replication but less active against the type 2 virus.

INTRODUCTION

A number of nucleoside analogues have recently been reported to be selective antiviral agents. We 3 and others have described the synthesis of 9-[(1,3-dihydroxy-2-propoxy)methyl]guanine (DHPG, 1) an

acyclic analogue of 2'-deoxyguanosine. DHPG is an exceptionally potent and selective inhibitor of herpes virus replication. In part, DHPG is selective because it is phosphorylated to its monophosphate by

a virus-specified thymidine kinase present only in infected cells. 6,7 The resulting monophosphate is then converted by cellular enzymes to the corresponding triphosphate of DHPG. 7 The nucleoside triphosphate analogue prevents herpes virus replication by inhibition of the virus-specified DNA polymerase. 8,9 Additional selectivity is realized because the nucleoside triphosphate is a better inhibitor of the viral than the host DNA polymerase. 8 The triphosphate also acts as an alternative substrate for the polymerase and once incorporated into the DNA leads to inhibition of chain elongation. 9 DHPG exhibits a broad spectrum of action being active against not only herpes simplex virus types 1 and 23,4a,6,10,11 but also cytomegalovirus, 4a,6,10,12 varicella-zoster, 4a,13 and Epstein-Barr virus. 4a,10,14

We have been synthesizing a number of analogues of DHPG in order to determine the effect of structural modifications on biological activity and now report the antiherpes activity of the carba analogue 2. The actual synthesis of 2 was previously reported without experimental details or biological data by Pandit and coworkers in 1972. The substitution of a methylene for the ether oxygen of nucleosides has many other precedents. For instance, the carbocyclic analogue of adenosine 3 (aristeromycin) was first synthesized in racemic form and later isolated as a natural product with antimicrobial activity. More recently the carbocyclic analogues of ara A (4) and 3-deazaadenosine (5) have been synthesized and shown to exhibit antiviral activity.

RESULTS AND DISCUSSION

Chemistry. Our synthesis of 2 (Scheme 1) is similar to that of Pandit's $1^{\overline{5}}$ and commences from diethyl malonate (6) which was alkylated with bromoacetaldehyde diethyl acetal (7) to give Diester 8 was reduced with lithium aluminum hydride to furnish diol 9.21 Benzylation of 9 by successive treatment with sodium hydride and then benzyl bromide afforded 10 in 64% distilled yield. Hydrolysis of 10 (p-toluenesulfonic acid, tetrahydrofuran/ water) followed by reduction (sodium borohydride) and distillation gave alcohol 11 in 81% yield. Reaction of 11 with p-toluenesulfonyl chloride furnished tosylate 12. In a related project, we found that primary tosylates did not react well with the sodium salt of guanine; therefore, 12 was treated first with sodium iodide and then the sodium salt of guanine to give the protected analogue 13 in 8% yield from 11. Cleavage of the benzyl ether functionalities of 13 by transfer hydrogenation²² (20% palladium hydroxide on carbon. cyclohexene, ethanol, reflux) afforded

9-[4-hydroxy-3-(hydroxymethyl)-1-butyl]guanine (2) in 79% yield.

The structures of both 2 and 13 were confirmed by examination of their carbon NMR spectra. The absorptions of the purine carbons proved that the side chain was indeed at $N^9.23$

Scheme 1

Antiviral activity of 2. Although 2 was nearly as potent in vitro as DHPG (1) against herpes simplex virus type 1 (HSV-1), it was substantially less effective against herpes simplex virus type 2 (HSV-2) (Table 1). Also the carba analogue 2 was seven-fold less effective than DHPG in inhibiting human cytomegalovirus (HCMV) replication.

In the mouse encephalitis model⁶ in which mice were challenged with HSV-2 (strain G), 2 was not effective at a dose of up to 20 mg/kg/day (Table 2). Additionally 2 did not prolong the mean survival time. The lack of activity of 2 as compared to DHPG in this in vivo model is consistent with the in vitro test data showing 2 to be less effective than 1 against HSV-2.

TABLE 1. Antiviral activities of carba-DHPG (2) and DHPG (1) in cell culture.

	ID ₅₀ (1	.м.) ^b
Virus ^a	2	1
HSV-1 (F)	0.5	.2
HSV-2 (G)	4.0	0.5
HCMV (AD 169)	45	7.0

^aThe strain is given in parentheses.

TABLE 2. Effects of oral treatment with carba-DHPG (2) and DHPG (1) on HSV-2 induced mortality in mice.

Drug	mg/kg ^a	Survivors/ Total	Survivor Increase ^b	Mean Survival Time (days)	Mean Survival Time Increase ^C
Saline		1/20 (5)d		9.4 <u>+</u> 1.9 ^e	
DHPG (1)	20	17/20 (85)	< .001	12.7 <u>+</u> 1.5	< .001
	10	16/20 (80)	< .001	12.3 <u>+</u> 1.0	< .001
	5	14/20 (70)	< .001	12.0 <u>+</u> 1.3	< .001
Carba-DHPG (2)	20	1/20 (5)	NS	9.7 <u>+</u> 1.8	NS
	10	4/20 (20)	NS	9.3 <u>+</u> 1.5	NS
	5	1/20 (5)	NS	9.3 <u>+</u> 1.6	NS

^aMice were infected intraperitoneally and an oral dose was administered once daily (at 24 h intervals) for 4 days starting 24 h after inoculation. Probability (Fisher exact test).

Determined by plaque assays in Vero (HSV) or MRC-5 (HCMV) cells.

CProbability (Mann-Whitney test).

dPercent survivors.

eStandard deviation.

TABLE 3. Effects of subcutaneous treatment with carba-DHPG (2) and DHPG (1) in a mouse HSV-1 intracerebral infection.

Drug	mg/kgª	Survivors/ Total	Survivor Increase ^b	Mean Survival Time (days)	Mean Survival Time Increase ^C
Saline		0/20 (0) ^d		6.2 <u>+</u> 1.4 ^e	
DHPG (1)	30	12/21 (57)	< .001	9.6 <u>+</u> 3.0	.001
	10	10/21 (48)	< .001	11.3 <u>+</u> 4.8	< .01
	3.0	10/21 (48)	< .001	8.7 <u>+</u> 2.6	< .01
	1.0	7/21 (33)	< .05	8.2 ± 3.4	NS
	0.3	2/22 (9)	NS	6.9 <u>+</u> 2.1	NS
Carba-DHPG (2)	100	2/17 (12)	NS	8.2 <u>+</u> 1.8	< ,001
	30	6/21 (29)	< .05	7.4 <u>+</u> 2.2	NS
	10	7/21 (33)	< .05	7.9 <u>+</u> 2.8	< .01

^aMice were infected intracerebrally and a subcutaneous dose of 2 or 1 was administered twice daily (at 12 h intervals) for 4 days starting 24 h post-infection.

Because carba-DHPG (2) and DHPG (1) are essentially equivalent against HSV-1 in vitro, we decided to also investigate their potency against a mouse intracerebral HSV-1 infection. ²⁴ In fact, 2 was effective, (Table 3) and at a dose of 10 mg/kg/day there were 7 out of 21 survivors in the carba-DHPG group compared to 10 out of 21 in the DHPG group. Possibly because of toxicity, 2 was not effective at 100 mg/kg/day.

Although carba-DHPG (2) is nearly as active as DHPG against HSV-1, the lower activity of 2 against HSV-2 and cytomegalovirus limits its potential utility.

bProbability (Fisher exact test).

CProbability (Mann-Whitney test).

dPercent survivors.

^eStandard deviation.

EXPERIMENTAL

Nuclear magnetic resonance spectra were recorded on a Varian EM-390 (¹H NMR, 90 MHz) and a Bruker WM-300 (¹H NMR, 300 MHz; ¹³C NMR, 75.453 MHz) and chemical shifts are reported in parts per million downfield from internal tetramethylsilane. Ultraviolet spectra were recorded on a Hewlett Packard 8450A spectometer. Spectroscopic data and elemental analyses were obtained by Syntex Analytical Research. All chromatographic purifications were carried out on silica gel. Melting points were determined on a hot-stage microscope and are corrected.

4-Benzyloxy-3-(benzyloxymethyl)-1-butanal diethyl acetal (10). To a stirred suspension of hexane prewashed NaH (13.4 g, 50%, 280 mmol) in DMF (400 mL) under N₂ was added 9^{21} (23.2 g, 121 mmol) as a solution in DMF (100 mL) over 15 min. After H₂ evolution ceased, benzyl bromide (64.9 mL, 264 mmol) was added over 0.5 h as a solution in DMF (100 mL). After 18 h, the solution was evaporated to dryness and the residue partition between ether and water. The organic phase was dried over MgSO₄ and evaporated to dryness. The residue was distilled (bp 186-190°C/1 torr) to give 28.8 g (64%) of 10 as a clear oil; ¹H NMR (300 MHz, CDCl₃) 7.30 (m, 10H, phenyl), 4.60 (t, J = 6 Hz, 1H, 0CH0), 4.48 (s, 4H, benzylic), 3.40-3.68 (m, 8H, CH₂0), 2.11 (heptet, J = 6 Hz, 1H, CH), 1.72 (t, J = 6 Hz, 2H, CH₂), 1.17 (t, J = 6 Hz, 6H, CH₃). Anal. Calcd for $C_{23}H_{32}O_4$ (372.50): C, 74.16; H, 8.66. Found: C, 74.29; H, 8.67.

4-Benzyloxy-3-benzyloxymethyl-1-butanol (11). A solution of 10 (28.2 g, 75.7 mmol) and -toluenesulfonic acid (0.35 g, 1.8 mmol) in THF (70 mL) and water (6 mL) was heated at reflux for 8 h then evaporated to dryness. The residue was dissolved in ethyl acetate washed with saturated NaHCO $_3$, dried over MgSO $_4$ and evaporated to dryness. A solution of the residue and NaBH $_4$ (2.36 g, 62.4 mmol) in methanol (70 mL) was stirred at room temperature for 0.5 h, the reaction was quenched with acetone and the solvent was evaporated. The resulting oil was dissolved in ethyl acetate, washed with 10% HCl,

saturated NaHCO $_3$ and water, dried over MgSO $_4$ and evaporated to dryness. The residue was distilled (bp 198-204°C/l torr) to give 18.4 g (81%) of 11 as a clear oil; ¹H NMR (90 MHz, CDCI $_3$) δ 7.30 (s, 10H, pheny1), 4.45 (s, 4H, benzylic), 3.30-3.80 (m, 6H, CH $_2$ 0), 2.86 (s, broad, 1H, 0H), 2.80 (septet, J = 6 Hz, 1H, CH), 1.64 (q, J = 6 Hz, 2H, CH $_2$). Anal. Calcd for C $_{19}$ H $_{24}$ O $_3$ (300.40): C, 75.97; H, 8.05. Found: C, 75.76; H, 8.13.

4-Benzyloxy-3-benzyloxymethyl-1-butyl tosylate (12). A solution of 11 (10.93 g, 36.3 mmol) and p-toluenesulfonyl chloride (9.46 g, 49.6 mmol) in pyridine (160 mL) was kept at 4°C for 18 h, then water (4 mL) was added. After 1 h, the solution was poured into ice water and extracted with ethyl acetate. The extract was washed with 5% HCl, saturated NaHCO₃ and brine, dried over MgSO₄ and evaporated to give 12 as a clear oil which was used directly in the next reaction. An analytical sample was prepared by chromatography (1:6 ethyl acetate/hexane); ¹H NMR (300 MHz, CDCl₃) & 7.74 (d, J = 8 Hz, 2H, tosylate), 7.22-7.37 (m, 12H, phenyl, tosylate), 4.42 (s, 4H, benzylic), 4.11 (t, J = 6 Hz, 2H, CH₂OTs), 3.43, 3.38 (ABX, J = 6 and 10 Hz, 4H, CH₂O), 2.42 (s, 3H, CH₃), 2.03 (heptet, J = 6 Hz, 1H, CH), 1.77 (q, J = 6 Hz, 2H, CH₂). Anal. Calcd for $C_{26}H_{30}O_{5}S$ (454.59): C, 68.70; H, 6.65; S, 7.05. Found: C, 68.60; H, 6.66; S, 7.07.

9-[(4-Benzyloxy-3-benzyloxymethyl)-1-butyl]guanine (13). A solution of 12 (12.0 g, 26.3 mmol) and NaI (7.91 g, 52.8 mmol) in DMF (100 mL) was stirred at room temperature for 1.5 h. In a separate flask, guanine (10.2 g, 67.2 mmol) plus NaH (2.71 g, 50%, 56.4 mmol; prewashed with hexane) in DMF (200 mL) was stirred at room temperature for 1 h. The two solutions were combined, heated at 90°C for 1.5 h and then evaporated to dryness. The residue was chromatographed (1:10 methanol/dichloromethane) and selected fractions recrystallized from ethanol to give 0.94 g (8%) of 13: mp 221-222°C; UV λ max (methanol) sh 275 nm (ϵ 9540), 253 (13700); 1 H NMR (300 MHz, Me₂SO-d₆) $^{\delta}$ 10.54 (s, broad, 1H, NH), 7.64 (s, 1H, H-8), 7.23-7.36 (m, 10H, phenyl), 6.40 (s, broad, 2H, NH₂), 4.42 (s, 4H, benzylic), 4.01 (t, J = 6 Hz, 2H, CH₂N), 3.35-3.50 (m, 4H, CH₂O), 1.83 (m,

3H, CH, CH₂); ¹³C NMR (75.453 MHz, Me₂SO-d₆) δ 156.72 (C-6), 153.34 (C-2), 151.05 (C-4), 138.41 (pheny1), 137.21 (C-8), 128.11, 127.30, 127.24 (pheny1), 116.57 (C-5), 72.05 (benzylic), 70.01 (CH₂O), 40.87 (CH₂N), 36.39 (CH), 29.01 (CH₂). Anal. Calcd for C₂₄H₂₇N₅O₃ (433.52): C, 66.49; H, 6.28; N, 16.16. Found: C, 66.37; H, 6.29; N, 16.15.

9-[(4-Hydroxy-3-hydroxymethy1)-1-buty1]guanine (2). A mixture of 13 (217 mg, 0.50 mmo1), 20% Pd(0H) $_2$ /C (220 mg), cyclohexene (5 mL) and ethano1 (15 mL) was heated at reflux for 18 h. The mixture was then diluted with 2:I methano1/water (200 mL) and filtered hot through celite. The filtrate was evaporated to dryness and the residue recrystallized from water to give 101 mg (79%) of 2: mp 273-275°C; λ max sh 273 nm (ϵ 10,050), 254 (14,100); 1 H NMR (Me $_2$ SO-d $_6$, 300 MHz) δ 10.59 (s, broad, IH, NH), 7.68 (s, 1H, H-8), 6.45 (s, 2H, NH $_2$), 4.44 (t, J = 5 Hz, 2H, 0H), 4.01 (t, J = 6 Hz, 2H, NCH $_2$), 3.40 (m, 4H, CH $_2$ 0), 1.72 (q, J = 6 Hz, 2H, CH $_2$), 1.46 (heptet, J = 6 Hz, 1H, CH); 13 C NMR (75.453 MHz, Me $_2$ SO-d $_6$) δ 156.68 (C-6), 153.13 (C-2), 151.00 (C-4), 137.48 (C-8), 116.41 (C-5), 79.01 (CH $_2$ 0), 40.97 (CH $_2$ N), 40.74 (CH), 28.66 (CH $_2$). Anal. Calcd for C $_{10}$ H $_{15}$ N $_{5}$ O $_3$ (253.26): C, 47.43; H, 5.97; N, 27.65. Found: C, 47.34; H, 6.00; N, 27.60.

Plaque Assays. Experiments were conducted with Vero cells infected with HSV-1 (F-strain) and HSV-2 (G-strain) or MRC-5 cells with HCMV (AD 169) and then treated with the nucleoside analogue as described previously. Fifty percent inhibitory doses (${\rm ID}_{50}$) are defined as doses causing a 50% reduction in plaque numbers compared to untreated controls.

Animal Studies. Swiss-Webster female mice (Simonsen Laboratories, Gilroy, Calif.), weighing approximately 20 g each, were infected intraperitoneally with 5 x 10⁴ plaque forming units of HSV-2 (strain G). This challenge was approximately equivalent to ten 50% lethal doses. Alternatively, the mice were infected intracerebrally with 250 plaque forming units of HSV-1 (strain Shealey). DHPG and 2 were administered subcutaneously once a day

for 4 days starting 24 h post-infection. Deaths were recorded for 21 days after infection.

Acknowledgement. We appreciate the assistance of Syntex
Analytical Research and especially Dr. M.L. Maddox, Mrs. J. Nelson and
Mrs. L. Kurz in obtaining and interpreting spectroscopic data.

REFERENCES

- 1. Contribution 197 from the Institute of Bio-Organic Chemistry, Syntex Research.
- (a) De Clercq, E. in "Targets for the Design of Antiviral Agents," De Clercq, E.; Walker, R.T., Eds., Plenum Press, New York, 1984, pp. 203-230. (b) Drach, J.C. in "Targets for the Design of Antiviral Agents," De Clercq, E.; Walker, R.T., Eds., Plenum Press, New York, 1984, pp. 231-258.
- 3. (a) Martin, J.C.; Dvorak, C.A.; Smee, D.F.; Matthews, T.R.; Verheyden, J.P.H. J. Med. Chem. 1983, 26, 759. (b) Verheyden, J.P.H.; Martin, J.C. US Patent 4,355,032, October 19, 1982.
- 4. (a) Field, A.K.; Davies, M.E.; DeWitt, C.; Perry, H.C.; Liou, R.; Germershausen, J.; Karkas, J.D.; Ashton, W.T.; Johnston, D.B.R.; Tolman, R.L. Proc. Natl. Acad. Sci. USA 1983, 80, 4139.
 (b) Ogilvie, K.K.; Cheriyan, U.O.; Radatus, B.K.; Smith, K.O.; Galloway, K.S.; Kennell, W.L. Can. J. Chem. 1982, 60, 3005.
 (c) Schaeffer, H.J. in "Nucleosides, Nucleotides and Their Biological Applications," Rideout, J.L.; Henry, D.W.; Beacham, L.M., Eds., Academic Press, New York, 1983, pp. 1-17.
- 5. The structural formulas of DHPG (1) and the related acyclic nucleoside analogues 2 and 13 have been depicted in a "ribose-like" conformation only to draw attention to the similarity in structure between these compounds and 2'-deoxy-nucleosides. In accordance with this representation, the two terminal carbons of the glycerol moiety are referred to as the 3' and 5' positions.
- 6. Smee, D.F.; Martin, J.C.; Verheyden, J.P.H.; Matthews, T.R. Antimicrob. Agents Chemother. 1983, 23, 676.
- Cheng, Y-C.; Grill, S.P.; Dutschman, G.E.; Nakayama, K.; Bastow, K.F. J. Biol. Chem. 1983, 258, 12460.
- 8. St. Clair, M.H.; Miller, W.H.; Miller, R.L.; Lambe, C.U.; Furman, P.A. Antimicrob. Agents Chemother. 1984, 25, 191.
- Frank, K.B.; Chiou, J-F.; Cheng, Y-C. <u>J. Biol. Chem.</u> 1984, <u>259</u>, 1566.
- Cheng, Y-C.; Huang, E-S.; Lin, J-C.; Mar, E-C.; Pagano, J.S.; Dutschman, G.E.; Grill, S.P. <u>Proc. Natl. Acad. Sci. USA</u> 1983, 80, 2767.

- 11. Smith, K.O.; Galloway, K.S.; Kennel, W.L.; Ogilvie, K.K.; Radatus, B.K. Antimicrob. Agents Chemother. 1982, 22, 55.
- 12. Mar, E-C.; Cheng, Y-C.; Huang, E-S. Antimicrob. Agents Chemother. 1983, 24, 518.
- 13. Bryson, Y.J. UCLA School of Medicine, Los Angeles, CA. Personal communication.
- 14. Lin, J-C.; Smith, M.C.; Pagano, J.S. J. Virology 1984, 50, 50.
- Pandit, U.K.; Grose, W.F.A.; Eggelte, T.A. <u>Synth. Comm.</u> 1972, 2, 345.
- 16. Shealy, Y.F.; Clayton, J.D. J. Am. Chem. Soc. 1966, 88, 3885.
- 17. Kusaka, T.; Yamamoto, H.; Shibata, M.; Muroi, M.; Kishi, T.; Mizuno, K. J. Antibiot. 1968, 21, 255.
- 18. (a) Vince, R.; Daluge, S. J. Med. Chem. 1977, 20, 612.
 (b) Shannon, W.M.; Westbrook, L.; Arnette, G.; Daluge, S.;
 Lee, H.; Vince, R. Antimicrob. Agents Chemother. 1983, 24, 538.
- (a) Montgomery, J.A.; Clayton, S.J.; Thomas, H.J.; Shannon, W.M.; Arnett, G.; Bodner, A.J.; Kion, I-K.; Cantoni, G.L.; Chiang, P.K. J. Med. Chem. 1982, 25, 626. (b) De Clercq, E.; Montgomery, J.A. Antiviral Research 1983, 3, 17.
- 20. Perkin, W.H.; Pink, H.S. J. Chem. Soc. 1925, 127, 91.
- 21. Raphael, R.A.; Roxburgh, C.M. J. Chem. Soc. 1955, 3405.
- 22. (a) Jackson, A.E.; Johnstone, R.A.W. Synthesis 1976, 685. (b) Anantharamaiah, G.M.; Sivanandaiah, K.M. J. Chem. Soc., Perkin Trans. 1 1977, 490.
- 23. Stothers, J.B. "Carbon-13 NMR Spectroscopy;" Academic Press; New York, 1972; pp. 469-473.
- Kern, E.R.; Richards, J.T.; Overall, J.C.; Glasgow, L.A. Antimicrob. Agents Chemother. 1978, 13, 53.

Received July 25, 1984.